



When the species is also a habitat: Comparing the predictively modelled distributions of *Lophelia pertusa* and the reef habitat it forms

Kerry L. Howell^{a,*}, Rebecca Holt^a, Inés Pulido Endrino^a, Heather Stewart^b

^a Marine Biology and Ecology Research Centre, Marine Institute at the University of Plymouth, Drake Circus, Plymouth PL3 5EE, UK

^b British Geological Survey, Murchison House, West Mains Road, Edinburgh EH9 3LA, UK

ARTICLE INFO

Article history:

Received 8 November 2010

Received in revised form 19 May 2011

Accepted 18 July 2011

Available online 12 August 2011

Keywords:

Deep-sea

Lophelia pertusa

Habitat mapping

Biotopes

Marine protected area

Maxent

ABSTRACT

Internationally there is political momentum to establish networks of marine protected areas for the conservation of threatened species and habitats. Practical implementation of such networks requires an understanding of the distribution of these species and habitats. Predictive modelling provides a method by which continuous distribution maps can be produced from limited sample data. This method is particularly useful in the deep sea where a number of biological communities have been identified as vulnerable ‘habitats’, including *Lophelia pertusa* reefs. Recent modelling efforts have focused on predicting the distribution of this species. However the species is widely distributed where as reef habitat is not. This study uses Maxent predictive modelling to investigate whether the distribution of the species acts as a suitable proxy for the reef habitat. Models of both species and habitat distribution across Hatton Bank and George Bligh Bank are constructed using multibeam bathymetry, interpreted substrate and geomorphology layers, and derived layers of bathymetric position index (BPI), rugosity, slope and aspect. Species and reef presence records were obtained from video observations. For both models performance is fair to excellent assessed using AUC and additional threshold dependant metrics. 7.17% of the study area is predicted as highly suitable for the species presence while only 0.56% is suitable for reef presence, using the sensitivity–specificity sum maximisation approach to determine the appropriate threshold. Substrate is the most important variable in the both models followed by geomorphology in the RD model and fine scale BPI in the SD model. The difference in the distributions of reef and species suggest that mapping efforts should focus on the habitat rather than the species at fine (100 m) scales.

© 2011 Elsevier Ltd. All rights reserved.

1. Introduction

The call for better spatial management of our marine environment is growing globally. Specifically, there is momentum for the establishment of networks of marine protected areas (MPAs) driven by global, European and (within the UK) national initiatives. One of the criteria by which MPAs are selected includes the protection of habitats and species that have been identified as rare, sensitive, functionally important, threatened and/or declining.

Within the NE Atlantic region the 1992 Convention for the Protection of the Marine Environment of the north-east Atlantic (OSPAR Convention) gives the OSPAR Commission a duty to develop means, consistent with international law, for establishing protective, conservation, restorative or precautionary measures related to specific species or habitats. A target date of 2010 has been set by OSPAR contracting parties to achieve “an ecologically

coherent network of well managed Marine Protected Areas” that serve (at least in part) to protect those habitats and species listed under Annex V of the Convention, on the OSPAR List of Threatened and/or Declining Species and Habitats.

At a European level the EU Habitats and Species Directive (92/43/EEC) requires the establishment of protected areas (Special Areas of Conservation – SACs) for habitats and species listed under Annex I and V respectively of the Directive, in areas of sea under the jurisdiction of member states (i.e. out to the 200 nm limit). In addition, the 2006 United Nations General Assembly Resolution 61/105 called “upon States to take action immediately, individually and through regional fisheries management organizations and arrangements, and consistent with the precautionary approach and ecosystem approaches, to sustainably manage fish stocks and protect vulnerable marine ecosystems (VMEs), including seamounts, hydrothermal vents and cold water corals, from destructive fishing practices”. This resolution has also ultimately resulted in the establishment of MPAs for the protection of specific species and habitats.

In order to establish MPAs for the protection of listed habitats and species there is a clear need to have a firm understanding of the distribution of those species and habitats (i.e. maps). The

* Corresponding author. Tel.: +44 (0)1752 584544.

E-mail addresses: kerry.howell@plymouth.ac.uk (K.L. Howell), hast@bgs.ac.uk (H. Stewart).

difficulties and expense of collecting species and habitat distribution data has led to the approach of using surrogates (Howell, 2010) and/or predictive species modelling techniques to provide maps of the distribution of vulnerable species (Bryan and Metaxas, 2007; Holmes et al., 2007; Embling et al., 2010). This approach is particularly useful for the deep-sea and high seas. Here, the vast area involved, sparse and highly localised data available, and distance from land, compound the problems encountered in shallow water settings. Within the deep-sea (high seas) ecosystem there are few actual species that are listed as of conservation concern under the legislation detailed above. Only commercially important fish species including orange roughy (*Hoplostethus atlanticus*), Portuguese dogfish (*Centroscymnus coelolepis*), and Leaf-scale gulper shark (*Centrophorus squamosus*), known to have undergone significant declines (ICES, 2008, 2010) are included. However a number of deep-sea habitats are listed. These habitats are predominantly biogenic in origin or are in fact biological assemblages, and include *Lophelia pertusa* reefs, coral gardens, sponge aggregations and sea-pen and burrowing megafauna communities, as well as other geogenic habitats such as carbonate mounds, seamounts, and oceanic ridges with hydrothermal vents/fields.

Recently, efforts have been made to model the distribution of the cold water coral *L. pertusa* from global to local scales, in order to identify areas of conservation importance (Davies et al., 2008; Dolan et al., 2008; Guinan et al., 2009b; Tittensor et al., 2009). *L. pertusa* is a widely distributed species and occurs as isolated colonies on boulders, cobbles, sand ripples, and even flat sea bed where some form of hard substrate is available for attachment (Wilson, 1979; Mortensen and Buhl-Mortensen, 2004a,b; Hovland, 2005). Its conservation importance stems from its reef forming capacity. *L. pertusa* can form large reefs and giant carbonate mounds up to 300 m high and several km in diameter (Roberts et al., 2006). Reef structures are highly biodiverse, possibly rivaling tropical coral reefs (Roberts et al., 2006). They may also have an important role as essential fish habitat but this is not yet clear (Husebø et al., 2002; Auster, 2005; Costello et al., 2005). *L. pertusa* only forms reefs under specific environmental conditions that are not yet fully understood but are controlled by the interplay between local hydrography and sedimentary dynamics (Thiem et al., 2006).

Given that the species is widely spread while the reef habitat has specific environmental requirements likely to result in a more confined distribution, to what extent does the distribution of *L. pertusa* species act as a proxy for the reef habitat? Mapping efforts that focus on the species rather than the habitat may produce maps of limited use to marine environmental managers if the distribution of the species is so broad as to indicate reef habitat is widely spread. Using the species distribution as a proxy for the habitat could provide a false impression of the extent of reef habitat and effect assessments of rarity and threat from human activities. The aim of this study is to use predictive modelling to investigate the difference in the distribution of the species and habitat and assess the implications of any difference to marine environmental management. Multibeam bathymetry, interpreted substrate and geomorphology layers, and derived layers of bathymetric position index (BPI), rugosity, slope and aspect are used as environmental input layers together with presence of *L. pertusa* reef and species. Here we conform to the definition of *L. pertusa* reef following Roberts et al. (2006) as biogenic structures formed by *L. pertusa* frameworks that alter sediment deposition, provide complex structural habitat and are subject to the processes of growth and (bio)erosion. However, presence data are derived from records of living *L. pertusa* reef only and not dead framework structures. The study focuses on Hatton and George Bligh Banks in the NE Atlantic.

2. Methods

2.1. Site description

Hatton Bank and George Bligh Bank are part of the Rockall Plateau (Hitchen, 2004), which is a large piece of continental crust that separated from the European continent during the early Cretaceous (Hauterivian–Cenomanian) when the North Atlantic was in the early stages of formation (Musgrove and Mitchener, 1996) (Fig. 1). Hatton Bank forms an elongate arc that stretches over 400 km and descends >2500 m below sea level into the Iceland Basin to the west and 1100 m below sea level into the Hatton Basin, (sometimes referred to as the Hatton–Rockall Basin), to the east. At its summit Hatton Bank lies less than 500 m below sea level. South of 59°N Hatton Bank is orientated approximately southwest–northeast, further north the orientation is more east–west. George Bligh Bank is broadly conical in shape and situated at the north-eastern end of the Rockall Plateau (and Hatton Basin). It rises to a summit at 450 m below sea level, and has a diameter of roughly 75 km.

2.2. Data collection

Collection of biological (video) data and low resolution multi-beam data from both Hatton Bank and George Bligh Bank were undertaken over a 1 month period (August–September) in 2005 using the commercial research vessel *S/V Kommandor Jack*. Further collection of biological data and collection of higher resolution multibeam data were undertaken over a 2 month period (August–October) in 2006 using the commercial research vessel *M/V Franklin*. Video sampling stations were selected during operations using multibeam bathymetry and backscatter data. Video tows were selected to cover a range of geomorphology, substrate types and water depths (Fig. 1).

Video data were collected using the Seatronics drop frame camera system. The system comprised an integrated DTS 6000 digital video telemetry system, which provided a real time video link to the surface, and a digital stills camera (5 mega pixel, Kongsberg OE14-208). In the 2005 surveys, the video stream from the viewing screen of the digital stills camera provided video data, in 2006 separate video (Kongsberg 14-366) and stills cameras were used. Cameras were mounted at an oblique angle (video: 24°; stills: 22° from the horizontal) to the sea bed to aid in species and habitat identification. Sensors monitored depth and altitude, and an Ultra Short Base Line (USBL) beacon provided accurate (to approximately 1 m) position data for the camera frame.

The system was deployed from the starboard side of the vessel. Video tows were between 250 and 1200 m long. For the majority of tows, vessel speed was approximately 0.5 knots (min 0.3 and max 0.7 knots), with most tows lasting between 0.5 and 1.5 h. The drop frame was towed in the water column between one and three metres (dependant on substrate type, topography and currents) above the sea bed. At the beginning of each tow, starting from when the sea floor became visible, a 2–3 min period was allowed before sampling, to enable the camera to stabilise before commencing the transect.

Videos were reviewed and the occurrence of *L. pertusa* colonies and *L. pertusa* reef habitat noted and linked to the navigational data from the USBL on the camera system, such that the location of each colony/habitat occurrence was recorded. Presence data for both species and habitat were then plotted in ArcGIS 9.3.

2.3. Multibeam

Multibeam data were acquired in 2005 on *S/V Kommandor Jack* using an EM120 (12 kHz; 191 beams) multibeam echosounder

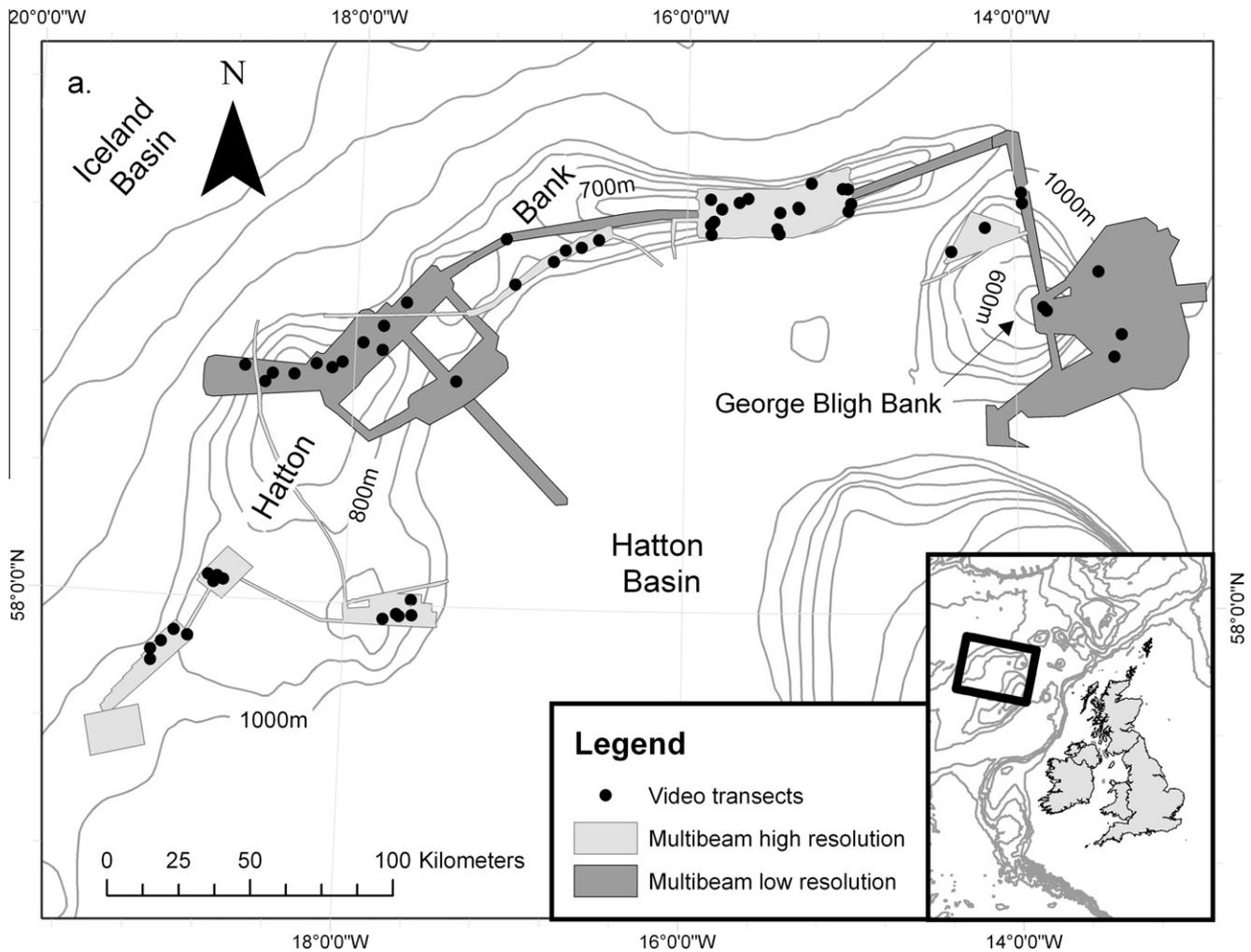


Fig. 1. The study area with sample details shown. Depth contours taken from the GEBCO digital atlas and are in 100 m isobaths down to 1000 m, thereafter in 500 m isobaths.

system which achieved good quality bathymetric data with marginal quality backscatter data. During the 2005 survey complete data coverage was achieved through 2500 m line spacing on Hatton Bank and 1500 m line spacing on the other survey areas. In 2006 multibeam data were acquired on *M/V Franklin* using an EM1002 (95 kHz; 111 beams) multibeam echosounder system which achieved excellent quality bathymetric and moderate quality backscatter data. During the 2006 survey complete data coverage was achieved through 650 m line spacing on Hatton Bank and 500 m line spacing on the other survey areas. It should be noted that weather and sea conditions adversely impacted on the backscatter data quality during both the 2005 and 2006 surveys. Positioning was accomplished using real-time Differential GPS (DGPS) systems. The C-Nav system was used in 2005 and the ARON 2000 system in 2006. All data acquisition systems took their time stamp from the primary DGPS which had a theoretical accuracy of better than 0.5 m. Sound velocity measurements were performed at regular intervals to account for hydrology effects during both surveys. Multibeam data were processed onboard ship and ashore by OSAE Ltd. in 2005 and Marin Mätteknik AB in 2006. Data were gridded at resolutions appropriate to the quality of the data (2005: 200 m grids; 2006: 25 m grids). Minimum waters depths encountered over Hatton Bank were -483 m, Lyonesse Seamount -507 m and George Bligh Bank -435 m. The maximum water depth of -1679 m was encountered on the eastern flank of George Bligh Bank as it descends into the Rockall Trough. As Maxent requires all environmental data to have the same geographic

bounds and cell size the 2006 multibeam dataset was regridded in ArcGIS 9.3 to a 200 m grid and merged with the 2005 multibeam dataset to produce a single bathymetry data layer, which was used to produce subsequent derived layers (see Section 2.4).

2.4. Derived layers

Additional environmental layers included in the model were layers derived from the multibeam bathymetry and were generated in ArcGIS using the spatial analyst and Benthic Terrain Modeller extensions. Derived layers included slope, rugosity (indicates the ratio of surface area to planar area), aspect (identifies the direction of the steepest slope) and bathymetric position index at broad and fine scale.

Bathymetric position index (BPI) is a measure of where a referenced location is relative to the locations surrounding it. Derived from an input bathymetric data set, a neighbourhood analysis function produces an output raster in which the output cell value at each location is a function of the input cell value and the values of the cells in a specified "neighborhood" surrounding that location. As bathymetric position is an inherently scale-dependent phenomenon (Weiss, 2001) both fine scale and broad scale BPI data sets are usually created, whereby the fine scale BPI layer is generated using a smaller analysis neighbourhood than the broad scale BPI layer. In the present study the default settings used in the Benthic Terrain Modeller extension to ArcGIS were applied to calculate coarse scale and fine scale BPI layers. These are, for broad scale BPI:

inner radius = 1, outer radius = 5, and fine scale: inner radius = 1, outer radius = 3. Positive cell values within a BPI data set denote features and regions that are higher than the surrounding area. Conversely negative cell values within a BPI data set denote features and regions that are lower than the surrounding area. BPI values near zero are either flat areas (where the slope is near zero) or areas of constant slope (where the slope of the point is significantly greater than zero) (Weiss, 2001).

2.5. Sea-bed substrate and geomorphological interpretations

All data were used to produce ArcGIS layers of sea-bed substrate and geomorphological features. For each digital stills image acquired a sea-bed sediment classification was assigned. These point classifications were used to ground-truth the multibeam echosounder and backscatter data allowing a complete sea-bed substrate interpretation to be created utilising all available data layers including derived layers. It should be noted that the backscatter quality was not suitable for accurate habitat differentiation as poor weather conditions and sea-state introduced noise which masked the more subtle geological variations of the sea floor. Following the sea-bed substrate classification, a geomorphological interpretation was created using standard geological terms and definitions. Geomorphology from the study area is described by 10 classes, with substrate described by 9 classes (Table 1).

The following eight environmental data layers were prepared in ArcGIS 9.3 for use in Maxent: continuous variables – bathymetry, slope, aspect, fine scale BPI, broad scale BPI, rugosity, categorical variables – substrate and geomorphology.

2.6. Maximum entropy modelling

Maximum entropy modelling was introduced as a general approach to presence only modelling of species distributions by Phillips et al. (2004, 2006). It has subsequently been shown to perform very well against other presence only models (Elith et al., 2006) with specific comparisons made between Maxent and environmental niche factor analysis (ENFA) applied to predictions of the global distribution of stony corals (Tittensor et al., 2009). Maxent estimates a target probability distribution by finding the probability distribution of maximum entropy subject to a set of constraints that represent our incomplete information about the target distribution (Phillips et al., 2006). Put simply and in the context of the present study Maxent allows the user to predict the distribution of a species/habitat in terms of probability of occurrence, by finding the distribution that agrees with everything known about the distribution of the species/habitat (given the environmental data that has been provided to the model), without making any assumptions about what is not known.

Table 1
Sea-bed substrate and geomorphology classes identified on Hatton Bank and George Bligh Bank.

Code	Substrate	Geomorphology
1	Gravelly sand	Scour
2	Gravel	Relatively flat lying sea bed
3	Bedrock	Pinnacle/mound
4	Gravelly muddy sand	Escarpment
5	Sand	Iceberg Ploughmarks
6	Mud	Furrow
7	Sandy gravel	Ridge crest
8	Muddy sand	Flank
9	Null	Channel
10	Sandy mud	Ridge
11	Null	Depression

Single models were constructed for *L. pertusa* species distribution (SD) and *L. pertusa* reef distribution (RD) using Maxent version 3.3.2, available for free download on <http://www.cs.princeton.edu/~schapire/maxent/>. Maxent was run with default settings: convergence threshold 10^{-5} and maximum number of iterations of 500, regularisation set to 1, that have been shown to achieve good performance (Phillips and Dudík, 2008) even with small size datasets. However, for the RD model following visual inspection of the response curves and subsequent trials with increased regularisation, the regularisation parameter was set to 2 to reduce over fitting of the model. Maxent results are given in a logistic system where values near 0 mean low probability of presence and values near 1 mean high probability of presence.

2.7. Model evaluation

The models generated were evaluated in two ways.

Firstly, threshold independent ROC (receiver operating characteristic) curves were used to measure how successful the prediction was using the area under the curve (AUC) (Fielding and Bell, 1997). As a result of the sampling method used (video transects), the presence data for both SD and more obviously for RD were spatially autocorrelated within transects. To attempt to account for this in the model evaluation process cross validation of the both models was performed manually rather than using the Maxent replicates setting. For the SD model 1479 presence records (reduced to 102 cells with presence records) were obtained from 43 transects. These data were partitioned such that approximately 25% of the transects (10 or 11) constituting ~25–30% of the presence records were omitted from model building and used as a test dataset. This process was repeated 10 times and average AUC and standard deviation of AUC across all 10 models was calculated. For the RD model the nature of the reef presence data was such that although there were nine cells with reef presence, in truth this amounted to observation of six complete reefs (as one reef occupied more than one cell). Therefore cross validation of the RD model was performed by splitting the presence data into groups corresponding to the six reefs observed and using these data to manually run the Maxent model six times, leaving out one ‘complete reef’ presence each time. Average AUC and standard deviation of AUC across all six models was calculated. However the small total number of presence samples available to the model suggests that cross validation may be inappropriate given that test data sets may consist of a single presence point.

Secondly, the model assessment indices: percent correctly classified (PCC), specificity and sensitivity (Fielding and Bell, 1997), were calculated using the Presence–Absence Model Evaluation library (Freeman, 2007) in R Development Core Team (2010)). These indices require that a threshold be used to convert the continuous Maxent probability of occurrence prediction to a binary prediction delimiting presence or absence. Determining the appropriate threshold for Maxent models is an interesting and ongoing area of research (Liu et al., 2005). In this study three possible thresholds were assessed for their use in producing a reliable binary output map (Table 2). The three selected for testing were from the group of metrics identified as ‘good’ by Liu et al. (2005). The effectiveness of each threshold was evaluated in R (R Development Core Team, 2010) using the Presence–Absence Model Evaluation library (Freeman, 2007) and model assessment indices listed previously. For SD and RD models the model build datasets were used together with absence data obtained from video analysis to assess the appropriateness of the thresholds. In addition thresholds were also assessed for each of the training and test model datasets and average performance of each threshold calculated. The most appropriate threshold was defined as that which resulted in constantly delivering the highest sensitivity score, since the precautionary

Table 2
Threshold dependent model performance metrics for SD and RD models for three different thresholding approaches.

SD models Threshold	Full model build data			Average training			Average test		
	PCC	Sensitivity	Specificity	PCC	Sensitivity	Specificity	PCC	Sensitivity	Specificity
Sensitivity–specificity equality	0.72	0.72	0.72	0.71 (0.08)	0.71 (0.08)	0.71 (0.08)	0.73 (0.12)	0.73 (0.12)	0.73 (0.12)
Sensitivity–specificity sum maximisation	0.67	0.92	0.58	0.70 (0.05)	0.81 (0.17)	0.68 (0.07)	0.80 (0.10)	0.72 (0.16)	0.82 (0.10)
ROC plot-based approach RD models	0.74	0.72	0.75	0.72 (0.03)	0.76 (0.13)	0.71 (0.03)	0.78 (0.10)	0.74 (0.14)	0.80 (0.10)
Sensitivity–specificity equality	0.90	0.94	0.90	0.79 (0.05)	0.79 (0.05)	0.79 (0.05)	0.93 (0.08)	0.96 (0.10)	0.93 (0.08)
Sensitivity–specificity sum maximisation	0.90	0.94	0.90	0.77 (0.14)	0.92 (0.11)	0.76 (0.14)	0.92 (0.11)	1.00 (0)	0.90 (0.13)
ROC plot-based approach	0.90	0.94	0.90	0.84 (0.11)	0.82 (0.07)	0.84 (0.11)	0.93 (0.08)	0.96 (0.10)	0.93 (0.08)

principle suggests that false positives are less of a concern than false absences. For this study the sensitivity–specificity sum maximisation approach where the sum of sensitivity and specificity is maximised Cantor et al. (1999), was selected (Table 2).

2.8. Assessment of variable importance within the models

Jackknife tests were undertaken to assess variable importance during model development by comparing the model gain (a measure of goodness of fit closely related to deviance) associated with models constructed with each variable omitted in turn, models constructed using individual variables only, and the full final model. Relative changes in gain between the full model and models constructed without one variable and with only one variable allow an assessment of relative importance of each variable to the final model build.

2.9. Quantification of species and habitat distribution

Binary maps for both SD and RD predicted distribution, produced using the sensitivity–specificity sum maximisation approach threshold obtained for the full models (SD = 0.2, RD = 0.25), were used to quantify the difference in area suitable for *L. pertusa* species and *L. pertusa* reef habitat in ArcGIS 9.3.

3. Results

3.1. Model evaluation

For all partitions of the occurrence data, for both the *L. pertusa* species distribution (SD) and *L. pertusa* reef distribution (RD) models, the AUC values achieved by the training–test data were better than random (Table 3) (AUC > 0.5). Full model AUC and mean training and test AUC for both SD and RD models could be rated a fair (0.7–0.8), good (0.8–0.9) or excellent (0.9–1). However, the consistently higher AUC value for the RD training–test models and the

higher AUCs for the full model (Table 3) indicates that the whole RD model can better discriminate between suitable and unsuitable distribution areas for *L. pertusa* reef than for *L. pertusa* species.

Threshold dependent model assessment indices for the sensitivity–specificity sum maximisation approach threshold also indicated that, when measured using PCC and sensitivity, the SD models performance was generally fair, while the RD models performance was generally excellent. Specificity scores for the SD models ranged between awful (<0.6), poor (0.6–0.7) and good (0.8–0.9) reflecting the decision to select a threshold to maximise sensitivity scores.

3.2. Importance of environmental variables within each model

Jackknife tests of variable importance revealed the models generated for both SD and RD relied heavily on the substrate variable both in terms of having the most useful information by itself and having the most information that was not present in the other variables (Fig. 2). Analysis of response curves created through construction of Maxent models using single variables illustrated the dependence of predicted suitability on substrate, with the bedrock, gravel and sandy gravel categories as most important in the SD model and bedrock as most important in the RD model (Fig. 3).

Within the RD model geomorphology was the second most important variable in terms of having the most useful information by itself, but was of limited importance to the SD model (Fig. 2). The geomorphological categories flank, pinnacle/mound, ridge, and escarpment were of most use. However, for both SD and RD models omission of the geomorphological variable resulted in the second largest drop in gain after depth and substrate respectively suggesting that the geomorphological layer contained information that was not present in the other variables (Fig. 2).

For both SD and RD models fine scale BPI and broad scale BPI were the next most important variables in terms of providing the highest gain when used in isolation to construct models (Fig. 2). As the information in one BPI layer was essentially also contained

Table 3
Area under the curve (AUC) scores for SD and RD models for full models and all partitions of the occurrence data into training–test datasets. Bold values highlight over all model performance.

SD Model	Training AUC	Test AUC	RD Model	Training AUC	Test AUC
Full model	0.964	0.808	Full model	0.998	0.940
<i>Cross validation models</i>					
1	0.839	0.695	1	0.924	0.792
2	0.567	0.774	2	0.909	0.870
3	0.856	0.744	3	0.946	0.912
4	0.634	0.884	4	0.854	1.000
5	0.797	0.721	5	0.876	1.000
6	0.815	0.957	6	0.876	1.000
7	0.822	0.854			
8	0.821	0.802			
9	0.828	0.627			
10	0.830	0.973			
Mean	0.781	0.803		0.897	0.929
Standard deviation	0.098	0.113		0.035	0.087

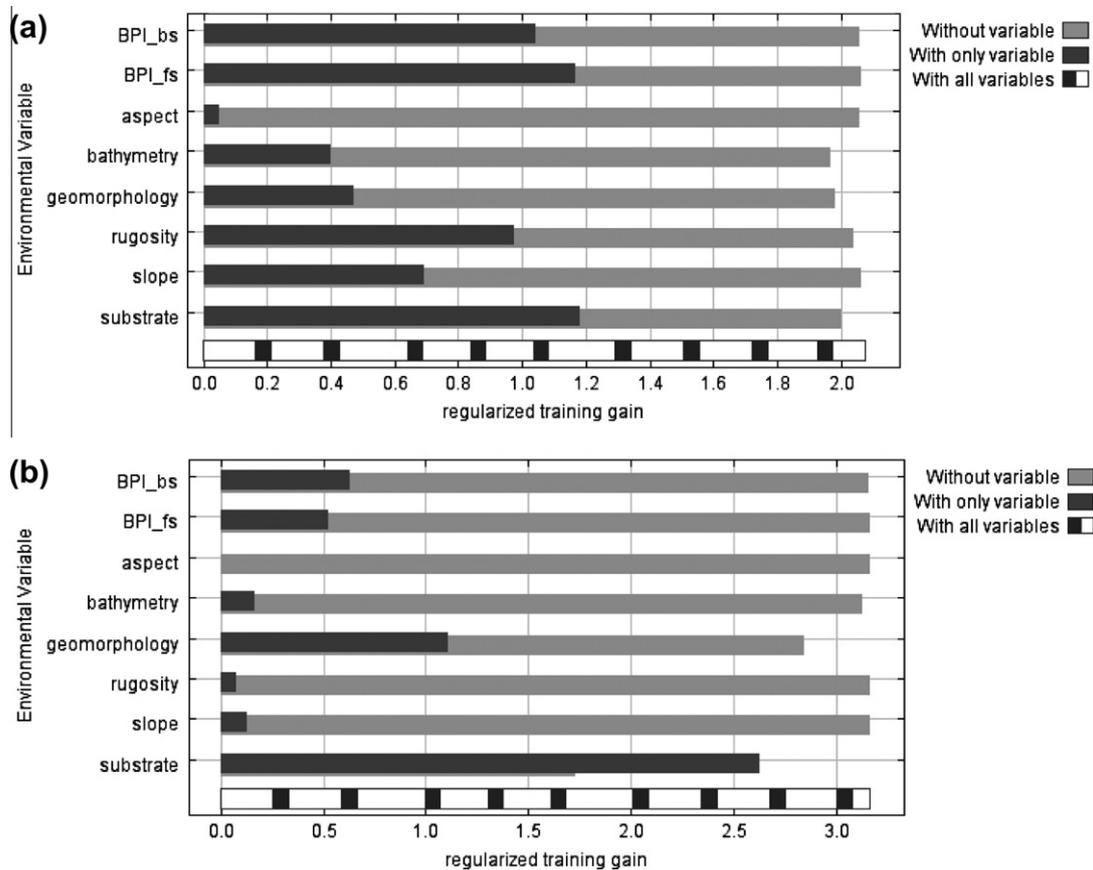


Fig. 2. Jackknife of regularised training gain for (a) *Lophelia pertusa* species and (b) *Lophelia pertusa* reef. “Without variable” – each variable is excluded in turn and a model created with the remaining variables; “With only variable” – model constructed using only one variable; “With all variables” – full model build.

in the other BPI layer, albeit at different resolution, in order to fully assess the importance of BPI to the SD and RD models, both models were rerun including only one BPI variable. However, in both the original models and the rerun models there was a negligible change in gain when BPI was omitted in Jackknife analyses. This suggests that BPI does not contain any information that was not present in other variables. Analysis of response curves illustrated subtle differences between models in the dependence of predicted suitability on both fine scale and broad scale BPI. Within the SD model both high negative values and high positive values were of greatest importance (Fig. 3). However within the RD model only high positive values were of greatest importance.

Within the SD model rugosity and slope were the next most important variables (Fig. 2). However, omission of the slope or rugosity variable in Jackknife tests resulted in a negligible drop in training gain. This suggests neither variable contained information that was not present in other variables. Analysis of response curves suggested the highest probability of occurrence of SD was achieved at rugosity of >1.01 and slopes of $>20^\circ$ (Fig. 3).

Jackknife tests of variable importance suggested that for both models bathymetry was of limited importance alone, however omission of the bathymetry variable from the models resulted in the third largest drop in training gain, suggesting the bathymetry variable contained information not present in other variables (Fig. 2). Heuristic estimates of relative contributions of the environmental variables to the Maxent model suggests that for SD and RD models bathymetry contributed 14.7% and 0.7% respectively. Analysis of response curves suggest the depths that resulted in the highest probability of SD presence were 500–900 m, however for RD the probability of presence was predicted to increase

with depth (Fig. 3), most likely a reflection of the limited depth range sampled.

Aspect was of least importance in both SD and RD models (Fig. 2) although interestingly probability of species occurrence was lowest at a bearing of 300° .

3.3. Potential distribution of *L. pertusa* species and reef

Binary maps (1-presence, 0-absence) of both species and habitat distribution show *L. pertusa* species is distributed over a broader area (7.17% of the map area) than *L. pertusa* reef habitat (0.56% of the map area) (Fig. 4).

4. Discussion

4.1. Species vs habitat distribution

On Hatton Bank and George Bligh Bank, the models identify a broader area of high suitability for the species than for reef (7.17% vs 0.56% of the total area respectively using the sensitivity-specificity sum maximisation threshold). Visual analysis of the spatial distribution of areas predicted as highly suitable for species and reef suggests that reef distribution is a highly restricted subset of species distribution.

Within the SD model the species was associated with bedrock, gravel and sandy gravel substrate categories, with no one geomorphological class of particular importance. These findings support what is currently known of the ecology of this species. *L. pertusa* has a cosmopolitan distribution (Zibrowius, 1980) and occurs as

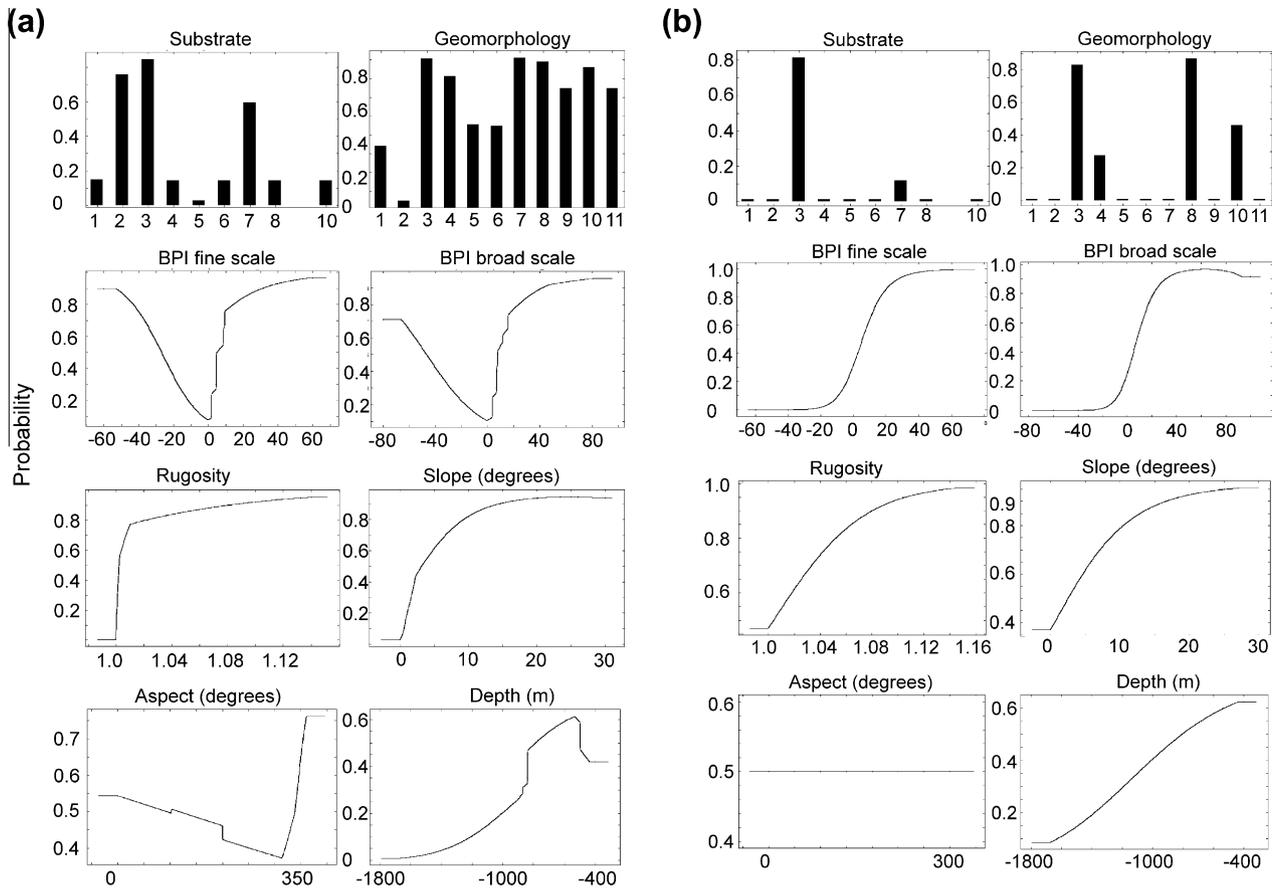


Fig. 3. Response curves generated from a model built using only the corresponding variable for (a) *Lophelia pertusa* species and (b) *Lophelia pertusa* reef. Y axis = probability of presence, X axis label given above each plot, or for substrate codes 1–10 and geomorphology codes 1–11, see Table 1.

individual colonies under a relatively broad range of conditions, from depths of 40–3400 m, temperatures of between 4 and 13 °C, salinities of 32–38‰ and across different oceans including the NE Atlantic, Barents Sea, the Mediterranean, and the Gulf of Mexico (Freiwald et al., 2004). *L. pertusa* is found on hard and mixed bottoms (Dons, 1944; Frederiksen et al., 1992) and in areas of fine sand where some form of hard substrate is present for initial attachment. Wilson (1979) suggested that suitable substrata for colony growth may be small e.g. mollusc shells, cobbles and boulders. It is not surprising then, to find that the species is likely to be found over a relatively wide area.

In comparison to the species, *L. pertusa* reef habitat is not widely distributed. The RD model indicates that reef habitat is only likely to be present over small areas on both Hatton Bank and George Bligh Bank. Within the wider NE Atlantic a limited number of large reef structures (mound regions) have been identified (see Wheeler et al. (2007) for a review). *L. pertusa* only forms reefs under a specific set of environmental conditions. The largest reefs occur in depths between 500 and 1200 m (Frederiksen et al., 1992; Wheeler et al., 2007) and may be associated with topographic features such as ridges (Sula Ridge), escarpments (Pelagia Mounds) and channels (Hovland Mounds) (Wheeler et al., 2007). Within the RD model reef habitat was clearly associated with bedrock substrate, and ridge, escarpment, flank and pinnacle/mound features.

While fundamental questions remain concerning the physical factors that are important in the development of reefs, recent research has highlighted the significance of hydrodynamic conditions in reef formation. Reef habitat forms in areas of enhanced turbidity, within a narrow density envelope, with high current velocities that prevent local sedimentation but provide enhanced

encounter-rates with food particles (Thiem et al., 2006; Mienis et al., 2007; Dullo et al., 2008). These conditions must be stable over long periods of time to allow reef development (Thiem et al., 2006). Inclusion of hydrographic data in the model would undoubtedly improve the model fit and predictive power, however fine scale oceanographic data are not widely available. Geomorphology acts as a surrogate for fine scale current speed. The relationship between reef habitat and geomorphological features such as ridges and escarpments identified by the model most likely reflects both the substrate and hydrodynamic requirements of reef habitat development.

4.2. Importance of environmental parameters to predictive modelling of *L. pertusa* species and habitat distribution

Within both the SD and RD models, substrate, geomorphology and BPI were the most important variables in terms of their importance to predicting species and habitat distribution, followed by rugosity and slope for SD and depth and slope for RD. These findings support those of Guinan et al. (2009a) who also found that at a slightly finer but comparable spatial scale (30 m multibeam grids) the most important variables in predicting *L. pertusa* species distribution using GARP modelling, were rugosity and slope. At finer resolution (0.5 m multibeam grids) Dolan et al. (2008) found BPI, structural complexity (fractal dimension) and orientation were important variables in using ENFA modelling. At coarser resolution (550 m grids) Guinan et al. (2009a) found in addition to rugosity and slope, that aspect was weakly important. Neither Dolan et al. (2008) nor Guinan et al. (2009a) had produced interpreted layers for substrate and geomorphology from multibeam bathymetry

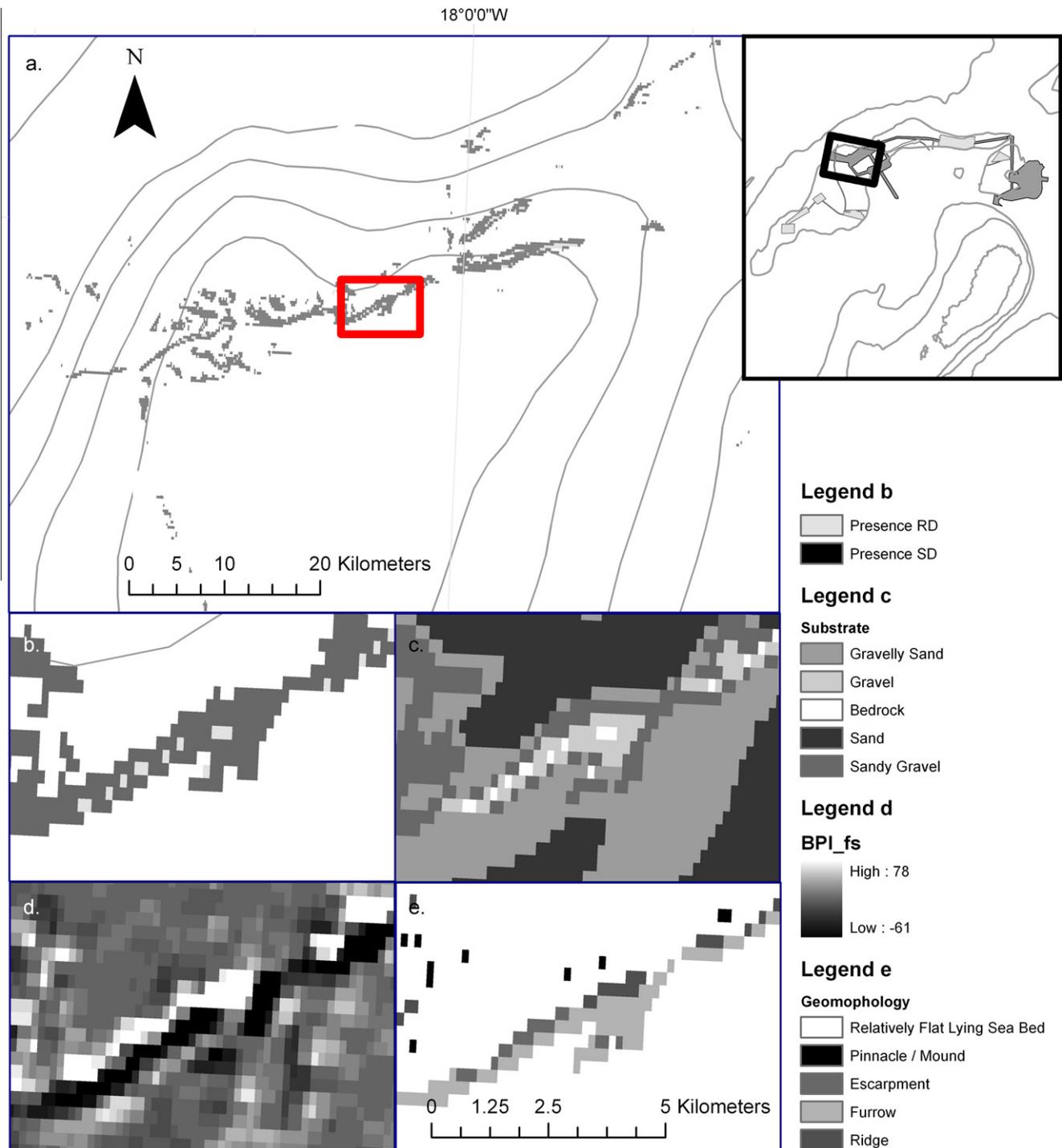


Fig. 4. Modelled distribution of *Lophelia pertusa* species and *Lophelia pertusa* reef (a) on a subsection of Hatton Bank (inset) and (b) on a small area identified in a. Maps c–e show the individual environmental layers from the same area as b, and illustrate the relationship between predicted presence areas and the two most important environmental variables in each model.

and backscatter and thus did not include these variables in their models. Following these studies Guinan et al. (2009b) concluded that coral abundance increases with increasing BPI, rugosity and slope.

Within the SD and RD models high positive BPI values were associated with high probability of occurrence suggesting both species and habitat are associated with raised features. High probability of species presence was also associated with high negative BPI values, suggesting the species may be associated with depressions as well as raised features. Probability of occurrence also

increased with increasing rugosity and slope to an asymptote (~ 1.16 and 30° respectively) suggesting values above this may not increase the probability of presence.

4.3. Implications for marine environmental management

The difference in area identified by the models as suitable for the species compared to the habitat has important implications for environmental management and the design of marine protected area networks (MPAs). As reef distribution is a restricted

subset of species distribution, calculations of habitat extent based on the species distribution (e.g. within a given countries EEZ or an area covered by a particular convention such as OSPAR in the NE Atlantic), will be gross overestimates, and will thus mask the relative rarity of the habitat. In addition, in the complex task of identifying suitable boundaries for MPAs for the purpose of conserving reef habitat, boundaries drawn on the basis of species distribution may fail to include the target habitat. It is therefore desirable, where possible, to focus on the distribution of the habitat over the species at least at fine scales. However, it would be unwise to consider *only* reef habitat distribution in conservation planning, since connectivity between reef areas is likely to be maintained by the wider species distribution. Isolated colonies on cobbles may well provide a mechanism for gene-flow between larger reefs. This is particularly important as gene flow occurring among subpopulations is moderate at best with high levels of inbreeding and self-recruitment (Le Goff-Vitry et al., 2004). Research is needed into issues of connectivity with respect to MPA planning.

The use of predictive species modelling in the deep-sea is a relatively new field. Recently models constructed at global and regional scales used broad-scale oceanographic data at cell sizes of 130 km, 1° and 0.25° to predict the distribution of *L. pertusa* species (Davies et al., 2008; Tittensor et al., 2009). These models identified the levels of nitrate silicate and phosphate, aragonite saturation, dissolved oxygen, and percent oxygen saturation as important in predicting the distribution of *L. pertusa*. The spatial resolution of the environmental data used in these models (in many cases data derived from model predictions) are inadequate to capture the fine-scale current regimes likely to determine the specific sites at which reefs are present (Guinan et al., 2009b). Therefore, while on global and regional scales a focus on modelling (and mapping) the distribution of the species is useful and appropriate for assisting in targeting and coordinating conservation efforts (Davies et al., 2008; Tittensor et al., 2009), the discrepancy between the areas of predicted presence of the species and the habitat in this study suggest that predictive modelling of habitat distribution at fine scales is more useful in terms of identifying areas of reef occurrence.

The importance of substrate, geomorphology, BPI, rugosity and slope to habitat distribution reflect the hydrodynamic conditions required for reef formation. This is important in terms of future mapping efforts. For *L. pertusa* reef, variables derived from multibeam bathymetry and interpreted backscatter data collectively act as suitable surrogates for those environmental factors, which are critical in determining reef distribution, but for which we generally lack fine-scale data. This suggests that it may be possible in future to undertake multibeam survey of appropriate resolution for large areas of the deep-sea and from that produce reasonable maps of reef distribution with limited ground truthing required. It also suggests that the final model produced here could be used to predict the distribution of reef habitat in other areas. However, the restricted depth and temperature range of the study area limits the final model to use in areas of similar environmental conditions.

What constitutes an appropriate resolution (multibeam grid size) for a given accuracy of predictive map requires further investigation if managers are to make informed decisions to balance predictive accuracy with survey cost. In addition the inclusion of interpreted substrate and geomorphology layers and their resulting importance in both SD and RD models suggests that these variables are particularly useful in providing good predictions. However, these interpretations take considerable time and skill to produce. In practical terms there may be a trade off between the time (and expense) taken to interpret such datasets and the gain in the accuracy of predicted distributions. Further research is needed into assessing such tradeoffs in the application of these methods.

4.4. Use of Maxent in predictive modelling of biological community distribution (biotope mapping)

The use of predictive modelling in marine community mapping is in its infancy (Kelly et al., 2001; Méléder et al., 2010). However, this technique has considerable benefits to offer to conservation efforts in the deep-sea where areas are vast, biological data are sparse and new survey is expensive. In shallow water areas remote sensing tools such as airborne and satellite imagery and aerial photography may be used to map the distribution of some habitats (Holmes et al., 2007). However, these tools rapidly reach their limits for sub-tidal surveys because of the absorption of visible radiation by water. At greater depths mapping is achieved using acoustic devices such as multibeam and sidescan sonar which are then ground truthed using video or other physical sampling methods (Brown et al., 2002; Huvenne et al., 2005; Brown and Blondel, 2009; Buhl-Mortensen et al., 2009). Although methods of mapping benthic assemblages vary, in general expert judgement is used to predict where assemblages will occur based on where they have been observed. This effectively amounts to predictive modelling using the mind. There is therefore a potential role for predictive modelling in biological assemblage (or biotope as defined by Dahl (1908)) mapping (Eastwood et al., 2006; Wilson et al., 2007). This study has demonstrated the potential use of the freely downloadable software Maxent to model the distribution of *L. pertusa* reef in benthic mapping efforts. This approach could be broadened and applied to other listed biogenic habitats/biological communities such as coral gardens, sponge aggregation, etc., as well as any other defined benthic assemblages (biotope).

Acknowledgements

The authors would like to acknowledge with thanks the scientists, officers and crew of S.V. Kommandor Jack and M.V. Franklin, the staff at Geotek and Marin Mätteknik A.B., the wider project partners J. Davies, C. Marshall, S. Mowles, L. Robinson, C. Jacobs, N. Golding, N. Coltman, and B. Narayanaswamy; also D. Bilton for helpful comments on the manuscript, C. Embling for useful discussions on thresholding and modelling in general, and two anonymous reviewers for their helpful comments. The collection of data used within was funded by the Department for Business, Enterprise and Regulatory Reform through Strategic Environmental Assessment 7 (formerly the Department for Trade and Industry) and the Department for Environment, Food and Rural Affairs through the their advisors the Joint Nature Conservation Committee and the offshore Special Areas for Conservation programme. Data analysis was partly funded by a mini grant awarded to K.L.H. from the Census of Seamounts, a joint Research Councils of the UK fellowship awarded to K.L.H, and University of Plymouth, and the ERASMUS student exchange programme. H.S. publishes with the permission of the Executive Director of the British Geological Survey (NERC).

References

- Auster, P.J., 2005. Are deep-water corals important habitats for fishes? In: Freiwald, A., Roberts, J.M. (Eds.), *Cold-water Corals and Ecosystems*. Springer, Berlin, Heidelberg, pp. 747–760.
- Brown, C.J., Blondel, P., 2009. Developments in the application of multibeam sonar backscatter. *Appl. Acoust.* 70, 1242–1247.
- Brown, C.J., Cooper, K.M., Meadows, W.J., Limpenny, D.S., Rees, H.L., 2002. Small scale mapping of sea-bed assemblages in the eastern English Channel using sidescan sonar and remote sampling techniques. *Estuar. Coast. Shelf Sci.* 54, 263–278.
- Bryan, T.L., Metaxas, A., 2007. Predicting suitable habitat for deep-water coral in the families Paragorgiidae and Primnoidae on the Atlantic and Pacific continental margins of North America. *Mar. Ecol. Prog. Ser.* 330, 113–126.

- Buhl-Mortensen, P., Dolan, M., Buhl-Mortensen, L., 2009. Prediction of benthic biotopes on a Norwegian offshore bank using a combination of multivariate analysis and GIS classification. *ICES J. Mar. Sci.* 66, 2026–2032.
- Cantor, S.B., Sun, C.C., Tortolero-Luna, G., Richards-Kortum, R., Follen, M., 1999. A comparison of C/B ratios from studies using receiver operating characteristic curve analysis. *J. Clin. Epidemiol.* 52, 885–892.
- Costello, M.J., McCrea, M., Freiwald, A., Lundalv, T., Jonsson, L., Brett, B.J., van Weering, T.C.E., de Haas, H., Roberts, J.M., Allen, D., 2005. Role of cold-water *Lophelia pertusa* coral reefs as fish habitat in the NE Atlantic. In: Freiwald, A., Roberts, J.M. (Eds.), *Cold-water Corals and Ecosystems*. Springer, Berlin, Heidelberg, pp. 771–805.
- Dahl, F., 1908. Grundsätze und grundbegriffe der bioecologischen forschung. *Zool. Anz.* 33, 349–353.
- Davies, A.J., Wisshak, M., Orr, J.C., Roberts, J.M., 2008. Predicting suitable habitat for the cold-water coral *Lophelia pertusa* (Scleractinia). *Deep-sea Res. Part I* 55, 1048–1062.
- Dolan, M.F.J., Grehan, A.J., Guinan, J.C., Brown, C., 2008. Modelling the distribution of cold-water corals in relation to bathymetric variables: adding spatial contact to deep-sea video. *Deep-sea Res. Part I* 55, 1564–1579.
- Dons, C., 1944. Norges korallrev. *K. Norske Vidensk. Selsk. Forh.* 16, 37–82.
- Dullo, C.W., Flögel, S., Rüggeberg, A., 2008. Cold-water coral growth in relation to the hydrography of the Celtic and Nordic European continental margin. *Mar. Ecol. Prog. Ser.* 371, 165–176.
- Eastwood, P.D., Souissi, S., Rogers, S.I., Coggan, R.A., Brown, C.J., 2006. Mapping seabed assemblages using comparative top-down and bottom-up classification approaches. *Can. J. Fish. Aquat. Sci.* 63, 1536–1548.
- Eliith, J., Graham, C.H., Anderson, R.P., Dudík, M., Ferrier, S., Guisan, A., Hijmans, R.J., et al., 2006. Novel methods improve prediction of species' distributions from occurrence data. *Ecography* 29, 129–151.
- Embling, C.B., Gillibrand, P.A., Gordona, J., Shrimpton, J., Stevick, P.T., Hammond, P.S., 2010. Using habitat models to identify suitable sites for marine protected areas for harbour porpoises (*Phocoena phocoena*). *Biol. Conserv.* 143, 267–279.
- Fielding, A.H., Bell, J.F., 1997. A review of methods for the assessment of prediction errors in conservation presence/absence models. *Environ. Conserv.* 24, 38–49.
- Frederiksen, R., Jensen, A., Westerberg, H., 1992. The distribution of the scleractinian coral *Lophelia pertusa* around the Faeroe Islands and the relation to internal tidal mixing. *Sarsia* 77, 157–171.
- Freeman, E., 2007. Presence Absence: An R Package for Presence–Absence Model Evaluation. USDA Forest Service, Rocky Mountain Research Station, 507 25th Street, Ogden, UT, USA.
- Freiwald, A., Fosså, J.H., Grehan, A., Koslow, T., Roberts, J.M., 2004. Coldwater Coral Reefs. UNEP-WCMC, Cambridge, UK.
- Guinan, J.C., Grehan, A.J., Wilson, M.F.J., Brown, C., 2009a. Quantifying relationships between video observations of cold-water coral and seafloor features in Rockall Trough, west of Ireland. *Mar. Ecol. Prog. Ser.* 375, 125–138.
- Guinan, J., Brown, C., Dolan, M.F.J., Grehan, A.J., 2009b. Ecological niche modelling of the distribution of cold-water coral habitat using underwater remote sensing data. *Ecol. Inform.* 4, 83–92.
- Hitchen, K., 2004. The geology of the UK Hatton-Rockall margin. *Mar. Petrol. Geol.* 21, 993–1012.
- Holmes, K.W., Van Niel, K.P., Kendrick, G.A., Radford, B., 2007. Probabilistic large-area mapping of seagrass species distributions. *Aquat. Conserv.: Mar. Freshwater Ecosyst.* 17, 385–407.
- Hovland, M., 2005. Pockmark-associated coral reefs at the Kristin field off Mid-Norway. In: Freiwald, A., Roberts, J.M. (Eds.), *Cold-water Corals and Ecosystems*. Springer, Berlin, Heidelberg, pp. 623–632.
- Howell, K.L., 2010. A benthic classification system to aid in the implementation of marine protected area networks in the deep/high seas of the NE Atlantic. *Biol. Conserv.* 143, 1041–1056.
- Husebø, Å., Nøttestad, L., Fosså, J.H., Furevik, D.M., Jørgensen, S.B., 2002. Distribution and abundance of fish in deep-sea coral habitats. *Hydrobiologia* 471, 91–99.
- Huvenne, V.A.L., Beyer, A., de Haas, H., Dekindt, K., Henriët, J.P., Kozachenko, M., Olu-Lé Roy, K., Wheeler, A.J., 2005. The seabed appearance of different coral bank provinces in the Porcupine Seabight, NE Atlantic, results from sidescan sonar and ROV seabed mapping. In: Freiwald, A., Roberts, J.M. (Eds.), *Cold-water Corals and Ecosystems*. Springer, Berlin, Heidelberg, pp. 535–569.
- ICES, 2008. Report of the Working Group on the Biology and Assessment of Deep-sea Fisheries Resources (WGDEEP). ICES Headquarters, Copenhagen.
- ICES, 2010. Report of the Working Group on Elasmobranch Fishes (WGEF), Horta, Portugal.
- Kelly, N.M., Fonseca, M., Whitfield, P., 2001. Predictive mapping for management and conservation of seagrass beds in North Carolina. *Aquat. Conserv.: Mar. Freshwater Ecosyst.* 11, 437–451.
- Le Goff-Vitry, M.C., Pybus, O.G., Rogers, A.D., 2004. Genetic structure of the deep-sea coral *Lophelia pertusa* in the northeast Atlantic revealed by microsatellites and internal transcribed spacer sequences. *Mol. Ecol.* 13, 537–549.
- Liu, C., Berry, P.M., Dawson, T.P., Pearson, R.G., 2005. Selecting thresholds of occurrence in the prediction of species distributions. *Ecography* 28, 385–393.
- Méléder, V., Populus, J., Guillaumont, B., Perrot, T., Mouquet, P., 2010. Predictive modelling of seabed habitats: case study of subtidal kelp forests on the coast of Brittany, France. *Mar. Biol.* 157, 1525–1541.
- Mienis, F., de Stigter, H.C., White, M., Duineveld, G., de Haas, H., vanWeering, T.C.E., 2007. Hydrodynamic controls on cold-water coral growth and carbonate-mound development at the SW and SE Rockall Trough Margin, NE Atlantic Ocean. *Deep-sea Res. Part I* 54, 1655–1674.
- Mortensen, P., Buhl-Mortensen, L., 2004a. Deep-water corals and their habitats in The Gully, a submarine canyon off Atlantic Canada. In: Freiwald, A., Roberts, J.M. (Eds.), *Cold-water Corals and Ecosystems*. Springer, Berlin, Heidelberg, pp. 247–277.
- Mortensen, P.B., Buhl-Mortensen, L., 2004b. Distribution of deep-water gorgonian corals in relation to benthic features in the Northeast Channel (Atlantic Canada). *Mar. Biol.* 144, 1223–1238.
- Musgrove, F.W., Mitchener, B., 1996. Analysis of the pre-Tertiary rifting history of the Rockall Trough. *Petrol. Geosci.* 2, 353–360.
- Phillips, S.J., Dudík, M., Schapire, R.E., 2006. Maximum entropy approach to species geographic distributions. *Ecol. Model.* 190, 231–259.
- Phillips, S.J., Dudík, M., 2008. Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. *Ecography* 31, 161–175.
- Phillips, S.J., Dudík, M., Schapire, R.E., 2004. A maximum entropy approach to species distribution modelling. In: Proceedings of the Twenty-first International Conference on Machine Learning. ACM Press, New York, p. 83.
- R Development Core Team, 2010. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, <<http://www.R-project.org>>.
- Roberts, J., Wheeler, A.J., Freiwald, A., 2006. Reefs of the deep: the biology and geology of cold-water coral ecosystems. *Science* 312, 543–547.
- Thiem, Ø., Ravagnan, E., Fosså, J.H., Bersten, J., 2006. Food supply mechanisms for coldwater corals along a continental shelf edge. *J. Mar. Syst.* 60, 207–219.
- Tittensor, D.P., Baco, A.R., Brewin, P.E., Clark, M.R., Consalvey, M., Hall-Spencer, J., Rowden, A.A., Schlacher, T., Stocks, K., Rogers, A.D., 2009. Predicting global habitat suitability for stony corals on seamounts. *J. Biogeogr.* 36, 1111–1128.
- Weiss, A.D., 2001. Topographic positions and landforms analysis. In: ESRI International User Conference, San Diego, USA.
- Wheeler, A.J., Beyer, A., Freiwald, A., de Haas, H., Huvenne, V.A.L., Kozachenko, M., Roy, K.O.L., Opperbecke, J., 2007. Morphology and environment of cold-water coral carbonate mounds on the NW European margin. *Int. J. Earth Sci.* 96, 37–56.
- Wilson, J.B., 1979. 'Patch' development of the deep-water coral *Lophelia pertusa* (L.) on Rockall Bank. *J. Mar. Biol. Assoc. UK* 59, 165–177.
- Wilson, M.F.J., O'Connell, B., Brown, C., Guinan, J.C., Grehan, A., 2007. Multiscale terrain analysis of multibeam bathymetry data for habitat mapping on the continental slope. *Mar. Geod.* 30, 3–35.
- Zibrowius, H., 1980. Les Scleractinaires de la Méditerranée et de l'Atlantique nord-oriental. *Mem. Inst. Oceanogr. (Monaco)* 11, 227.